

Homoploid Hybrid Speciation in an Extreme Habitat

Zachariah Gompert,¹ James A. Fordyce,² Matthew L. Forister,³ Arthur M. Shapiro,⁴ Chris C. Nice^{1*}

According to theory, homoploid hybrid speciation, which is hybrid speciation without a change in chromosome number, is facilitated by adaptation to a novel or extreme habitat. Using molecular and ecological data, we found that the alpine-adapted butterflies in the genus *Lycaeides* are the product of hybrid speciation. The alpine populations possess a mosaic genome derived from both *L. melissa* and *L. idas* and are differentiated from and younger than their putative parental species. As predicted, adaptive traits may allow for persistence in the environmentally extreme alpine habitat and reproductively isolate these populations from their parental species.

Homoploid hybrid speciation is characterized by hybridization between parental species that results in a derivative hybrid species without a change in chromosome number (1–6). A growing list of possible examples, such as African cichlids (7), cyprinid fishes (8), *Rhagoletis* fruit flies (9), *Heliconius* butterflies (10), and swallowtail butterflies (11), suggest that homoploid hybrid speciation in animals may be more common than previously thought. Models predict that ecological isolation spurs homoploid hybrid speciation, especially when the hybrids invade novel or extreme habitats (12). Colonization of a novel habitat by an incipient hybrid species may allow it to avoid introgression and competition with the parental species (4, 12). Although these predictions have been borne out in plants (13), no examples of homoploid hybrid speciation in animals have involved adaptation to a novel habitat, although a switch to a novel host plant species has been documented (9).

The ecologically, morphologically, and behaviorally distinct species *L. melissa* and *L. idas* (14–17) have come into secondary contact in the Sierra Nevada of western North America (18) (Fig. 1). *Lycaeides melissa* populations occur in Great Basin habitats on the east side of the Sierra Nevada, whereas *L. idas* populations occupy wet meadows at mid-elevation on the west slope of these mountains. Unnamed populations of *Lycaeides* occur in the alpine habitat above the tree line of the Sierra Nevada, an environmentally extreme habitat not occupied by *L. melissa* or *L. idas*. The alpine habitat is characterized by a short growing season and severe fluctuations in ambient temperature and relative humidity on a daily and seasonal basis (19). These alpine butterflies have male genitalia that are interme-

diated in size and shape compared with those of *L. melissa* and *L. idas* (18), with wing pattern elements that are qualitatively similar to those of *L. melissa* (14). However, analysis of mitochondrial DNA variation shows that the alpine populations' haplotypes share a more recent

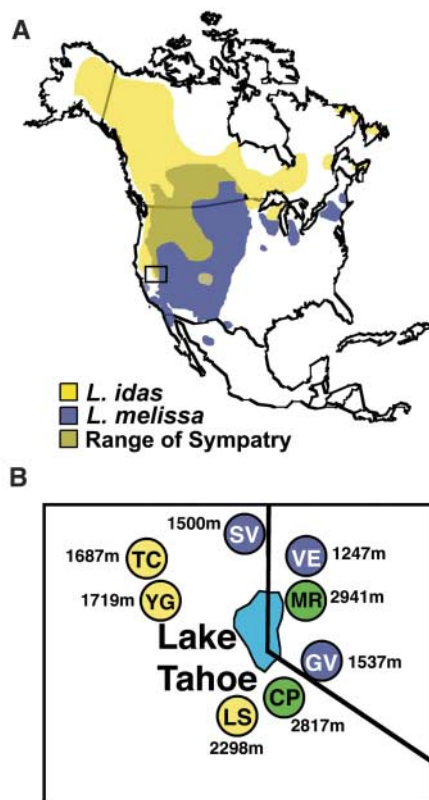


Fig. 1. Approximate range of *L. melissa* and *L. idas* in North America (A) and sampling localities for this study (B). The range map is based on Nabokov (23), Stanford and Opler (24), and Scott (25). *Lycaeides idas* is shown in yellow, *L. melissa* is shown in blue, and regions of sympatry are shown in gray. The box denotes the focal region for this study. Sampling localities for *L. idas*, *L. melissa*, and alpine *Lycaeides* are shown in yellow, blue, and green, respectively. SV, Sierraville; VE, Verdi; MR, Mt. Rose; GV, Gardnerville; CP, Carson Pass; LS, Leek Springs; YG, Yuba Gap; TC, Trap Creek.

common ancestor with haplotypes of *L. idas* than those of *L. melissa* (fig. S1) (20). These discordant patterns suggest that hybridization may have played a role in the evolutionary history of alpine *Lycaeides* populations.

If the alpine *Lycaeides* populations are a hybrid species, they should possess a genome that is a blend of alleles derived from both *L. melissa* and *L. idas*. We tested this using a large multilocus genomic data set consisting of 128 amplified fragment length polymorphism (AFLP) markers, three microsatellite markers (*Msat201*, *Msat4*, and *MsatZ12-1*), and sequence data from three nuclear genes (*Nuc1*, *Nuc3*, and *Ej1a*) and two mitochondrial genes (*COI* and *COII*) (20). To assess the overall genomic composition of the alpine *Lycaeides* populations, we used the Bayesian program STRUCTURE version 2.1 to cluster *L. melissa*, *L. idas*, and alpine individuals on the basis of their multilocus genotypes (20, 21) under the assumption that the data represented two separate populations ($K = 2$). Individuals from *L. melissa* and *L. idas* clustered to different groups with high probability, whereas alpine *Lycaeides* individuals were assigned to both groups with moderate probability (Fig. 2A and table S1). This pattern is inconsistent with a bifurcating mode of speciation, in which alpine butterflies originated from a single parental species, and suggests that the alpine genome is a mosaic of the two species. In further support of this hypothesis, five AFLP fragments were shared between *L. melissa* and *L. idas* to the exclusion of the

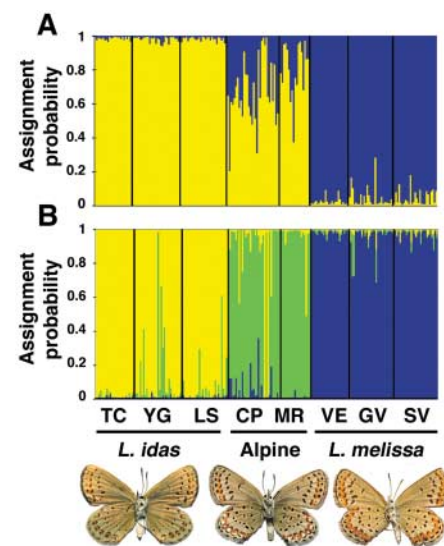


Fig. 2. Bar plots showing Bayesian assignment probabilities from the software STRUCTURE 2.1 (21) for two (A) and three (B) clusters (20). Each vertical bar corresponds to one individual. The proportion of each bar that is yellow, blue, and green represents an individual's assignment probability to clusters one, two, and three, respectively. See table S1 for mean population assignment probabilities. Location abbreviations are as in Fig. 1.

¹Department of Biology, Population and Conservation Biology Program, Texas State University, San Marcos, TX 78666, USA. ²Department of Ecology and Evolutionary Biology, University of Tennessee, Knoxville, TN 37996, USA. ³Department of Natural Resources and Environmental Science, University of Nevada, Reno, NV 89512, USA. ⁴Section of Evolution and Ecology, University of California, Davis, CA 95616, USA.

*To whom correspondence should be addressed. E-mail: ccnice@txstate.edu

alpine populations, whereas the alpine populations shared 12 unique alleles with *L. melissa* and 16 unique alleles with *L. idas*. Additionally, in *L. melissa* and *L. idas*, different alleles were fixed at the *Nuc1* locus (fig. S2 and table S1), whereas the alpine populations shared three *Nuc1* alleles with *L. melissa* and three *Nuc1* alleles with *L. idas*.

Although the mosaic genome of the alpine populations is consistent with homoploid hybrid speciation, a similar pattern could arise if the alpine populations have continuous gene flow with *L. melissa* and/or *L. idas*. If so, the alpine populations would not be genetically differentiated from *L. melissa* and *L. idas*, and there would be evidence of gene flow with these species. When STRUCTURE (21) was run under the assumption that the data represented three separate populations ($K = 3$), *L. melissa* and *L. idas* individuals were still assigned to their respective clusters, but the alpine *Lycaeides* individuals were assigned to a distinct, third cluster (Fig. 2B and table S1) (20). Additionally, alpine populations were fixed for unique alleles at the mitochondrial genes *COI* and *COII*, as well as the nuclear gene *Nuc3* (figs. S1 and S3 and table S1). These data and examination of pairwise F_{ST} (20) (table S2) suggest that alpine populations are differentiated from *L. melissa* and *L. idas*. We did not detect excess heterozygosity or deviations from linkage equilibrium for any microsatellite markers or nuclear gene sequences (20); such deviations would indicate ongoing gene flow between the alpine populations and either *L. melissa* or *L. idas*. We used the program NewHybrids, which uses the Bayesian assignment algorithm of Anderson and Thompson (22) to assess the probability that gene flow occurs between *L. melissa* and/or *L. idas* and the alpine populations (20). No individuals were identified that could be considered F_1 's produced from crosses between *L. melissa* or *L. idas* and the alpine populations

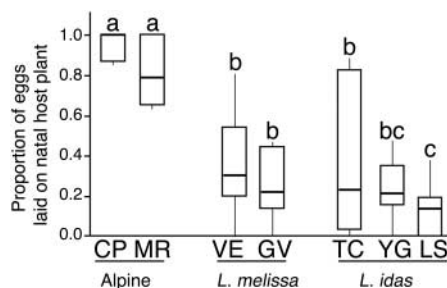


Fig. 3. Natal host plant fidelity from seven focal populations. Box plots show the median proportion of eggs laid on the natal host plant for each population. Kruskal-Wallis test indicates significant differences in natal host plant preference among populations ($T = 46.67$; $P < 0.0001$). Different letters (a, b, and c) indicate differences in strength of preference for natal host among populations ($\alpha < 0.05$). Natal host plants are listed in table S1. Location abbreviations are as in Fig. 1.

(fig. S4). Thus, we concluded that the alpine populations are genetically differentiated from *L. melissa* or *L. idas* and are not exchanging genes with either.

The genetic patterns we documented could have occurred if *L. melissa*, *L. idas*, and the alpine populations all developed rapidly from a single ancestral species distributed along a geographic cline with the alpine populations originating from the center of the cline. This scenario is unlikely for several reasons. Phylogeographic data suggest that the current distribution of *L. melissa* and *L. idas* is the result of post-Pleistocene range expansion and secondary contact and thus does not reflect the distribution of the ancestor of these species (18). The alpine populations also have a more recent origin than either *L. melissa* or *L. idas*. We calculated a coalescent-based estimate of the time to the most recent common ancestor (TMRCA) for mitochondrial variation for each of the three putative species: the alpine populations, *L. melissa*, and *L. idas* (20). The estimated TMRCA for the alpine populations, 442,579 years before present (yr B.P.), is substantially younger than that of either *L. melissa* or *L. idas*, 1,902,995 and 1,267,885 yr B.P., respectively (20). Furthermore, pairwise estimates of τ (species divergence time multiplied by mutation rate) based on nuclear and mitochondrial sequence data were approximately four times greater for the divergence of *L. melissa* and *L. idas* [0.006576, 95% confidence interval (95% CI) 0.002823 to 0.009855] than for the divergence of the alpine populations and either *L. melissa* (0.001318, 95% CI 0.000638 to 0.002233) or *L. idas* (0.001468, 95% CI 0.000763 to 0.002454) (20) (fig. S5). Thus, *L. melissa*, *L. idas*, and the alpine populations did not arise rapidly from a single ancestral species.

Homoploid hybrid speciation is more likely when a hybrid species colonizes a novel habitat (5, 6, 12, 13), such as the alpine habitat occupied by *Lycaeides*. Reproductive isolation between the hybrid and the parental taxa may be maintained by behavioral and ecological adaptations to the alpine habitat specifically associated with the alpine host plant. Females from the alpine populations have near-perfect host fidelity for their host plant, the perennial alpine endemic, *Astragalus whitneyi* (Fig. 3) (20). Indeed, alpine females have stronger host fidelity than has been recorded in other *Lycaeides* populations. Because males and females of *Lycaeides* locate mates and copulate on or near their larval host plants (15), strong fidelity with *A. whitneyi* may serve as a strong prezygotic barrier to gene flow between alpine *Lycaeides* populations and their putative parental species. In addition, host fidelity is coupled with a unique lack of egg adhesion in the alpine populations. Whereas *Lycaeides* females from nonalpine populations “glue” their eggs to their host plant when they oviposit, the alpine populations’ eggs fall off the plant after oviposition and remain

near the site of new plant growth in the following spring (16). Because *Lycaeides* overwinter as diapausing eggs (16) and the senesced aboveground biomass of the alpine host plant is blown away by strong winds in the winter (16), any eggs attached to the alpine host plant would be carried far from the site of new host plant growth, resulting in likely death of neonate larvae. Any females from nonalpine populations that oviposit on the alpine host plant would suffer a major reduction in fitness. Together, these alpine-associated adaptive traits—strong host fidelity for an alpine endemic host plant and the loss of egg adhesion—may act as an effective ecological barrier to gene flow.

Two other mechanisms may also contribute to reproductive isolation. Color pattern differences on the underside of the wings operate as species recognition cues, isolating the alpine populations from *L. idas* (14). Differences in male genital morphology have also been documented (18) and may operate in a similar manner to limit gene flow, although this was not explicitly tested in this study. Thus, morphological characters and adaptation to an extreme, novel habitat may create reproductive isolation between the hybrid species and its parental species.

References and Notes

1. J. A. Coyne, H. A. Orr, *Speciation* (Sinauer Associates, Sunderland, MA, 2004).
2. V. Grant, *Plant Speciation* (Columbia Univ. Press, New York, 1981).
3. M. L. Arnold, *Natural Hybridization and Evolution* (Oxford Univ. Press, Oxford, 1997).
4. T. E. Dowling, C. L. Secor, *Annu. Rev. Ecol. Syst.* **28**, 593 (1997).
5. L. H. Rieseberg, *Annu. Rev. Ecol. Syst.* **28**, 359 (1997).
6. B. L. Gross, L. H. Rieseberg, *J. Hered.* **96**, 241 (2005).
7. R. Schelly, W. Salzburger, S. Koblmüller, N. Duftner, C. Sturmhuber, *Mol. Phylogenet. Evol.* **38**, 426 (2006).
8. B. D. Demarais, W. L. Minckley, *Copeia* **1992**, 697 (1992).
9. D. Schwarz, B. M. Matta, N. L. Shakir-Butterli, B. A. McPherson, *Nature* **436**, 546 (2005).
10. J. Mavarez *et al.*, *Nature* **441**, 868 (2006).
11. J. M. Scriber, G. J. Ordling, *Entomol. Exp. Appl.* **115**, 247 (2005).
12. C. A. Buerkle, R. J. Morris, M. A. Asmussen, L. H. Rieseberg, *Heredity* **84**, 441 (2000).
13. L. H. Rieseberg *et al.*, *Science* **301**, 1211 (2003).
14. J. A. Fordyce, C. C. Nice, M. L. Forister, A. M. Shapiro, *J. Evol. Biol.* **15**, 871 (2002).
15. C. C. Nice, J. A. Fordyce, A. M. Shapiro, R. French-Constant, *Ecol. Entomol.* **27**, 702 (2002).
16. J. A. Fordyce, C. C. Nice, *Ecol. Lett.* **6**, 23 (2003).
17. M. L. Forister, J. A. Fordyce, C. C. Nice, Z. Gompert, A. M. Shapiro, *Ann. Entomol. Soc. Am.* **99**, 933 (2006).
18. C. C. Nice, N. Anthony, G. Gelembiuk, D. Raterman, R. French-Constant, *Mol. Ecol.* **14**, 1741 (2005).
19. M. V. Lomolino, B. R. Riddle, J. H. Brown, *Biogeography* (Sinauer Associates, Sunderland, MA, ed. 3, 2006).
20. Materials and methods are available as supporting material on Science Online.
21. J. K. Pritchard, M. Stephens, P. Donnelly, *Genetics* **155**, 945 (2000).
22. E. C. Anderson, E. A. Thompson, *Genetics* **160**, 1217 (2002).
23. V. Nabokov, *Bull. Mus. Comp. Zool.* **101**, 479 (1949).
24. R. E. Stanford, P. A. Opler, *Atlas of Western USA Butterflies, Including Adjacent Parts of Canada and Mexico* (Self-published, Fort Collins, CO, 1996).
25. J. A. Scott, *The Butterflies of North America, a Natural History and Field Guide* (Stanford Univ. Press, Stanford, CA, 1986).

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lab. GenBank accession numbers for sequence data are EF 090312-EF090397.

Tables S1 to S4
References

Supporting Online Material

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Materials and Methods
SOM Text
Figs. S1 to S10

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A Giant European Dinosaur and a New Sauropod Clade

Rafael Royo-Torres,* Alberto Cobos, Luis Alcalá

Fossils of a giant sauropod dinosaur, *Turiasaurus riodevensis*, have been recovered from terrestrial deposits of the Villar del Arzobispo Formation (Jurassic-Cretaceous boundary) of Riodeva (Teruel Province, Spain). Its humerus length (1790 millimeters) and estimated mass (40 to 48 metric tons) indicate that it may have been the most massive terrestrial animal in Europe and one of the largest in the world. Phylogenetic analysis indicates that the fossil represents a member of a hitherto unrecognized group of primitive European eusauropods that evolved in the Jurassic.

Most gigantic sauropods are neosauropods and have been found in the New World [such as *Seismosaurus* (1) and *Sauroposeidon* (2) in North America and *Argentinosaurus* (3) and *Puertasaurus* in South America (4)] or Africa [such as *Paralititan* (5) and *Brachiosaurus* (6)]. Here we describe a new giant European sauropod from Riodeva (Spain) as a new taxon, *Turiasaurus riodevensis* gen. et sp. nov., in Tithonian-Berriasian-aged deposits of the Villar del Arzobispo Formation (7) (figs. S1 and S2). The Barrihonda–El Humero site, where the new gigantic sauropod was found, has also yielded theropod teeth, postcranial remains of stegosaurs, and isolated elements of ornithomorphs, as well as fishes, turtles, and crocodylomorphs. The only other sauropod specimens reported from Riodeva were isolated elements from Pino de Jarque 2 and La Cautiva 2 (fig. S2): an ilium of a Diplodocidae indet (8) and a proximal caudal vertebra of a basal eusauropod (9).

Etymology. *Turiasaurus*, from Turia (word used since the 12th century from which Teruel derives) and *sauros* (Greek word, lizard); *riodevensis*, from Riodeva (village where the fossil site is located).

Holotype. An articulated left forelimb (Fig. 1, A to G) including humerus, radius, proximally incomplete ulna, carpal, five metacarpals, and seven phalanges (specimen numbers CPT-1195 to CPT-1210, housed in the Museo de la Fundación Conjunto Paleontológico de Teruel-Dinópolis, Teruel, Aragón, Spain).

Fundación Conjunto Paleontológico de Teruel-Dinópolis. Avenida de Sagunto, E-44002 Teruel, Spain.

*To whom correspondence should be addressed. E-mail: royo@dinopolis.com

Paratype. Remains attributed to the same individual, found close to each other in an area of 280 m² (fig. S3), consisting of skull frag-

ments, eight teeth (Fig. 2), six cervical vertebrae (Fig. 1K) with ribs (Fig. 1, L and M) (probably cervicals 3 to 8), two proximal dorsal vertebrae, a middle dorsal vertebra (Fig. 1I), fragments of other dorsal vertebrae, eight dorsal ribs (five incomplete), a partial sacrum, two distal caudal vertebrae (Fig. 1T), a proximal fragment of the left scapula, a left sternal plate (Fig. 1J), a distal fragment of the left femur, a proximal fragment of the left tibia (Fig. 1, N and O), a distally incomplete left fibula (Fig. 1H), a complete left astragalus (Fig. 1, P and Q), two left pedal phalanges, and an incomplete right astragalus and pes (Fig. 1, R and S) (CPT-1211 to CPT-1261), housed in the Museo de la Fundación Conjunto Paleontológico de Teruel-Dinópolis, Teruel, Spain).

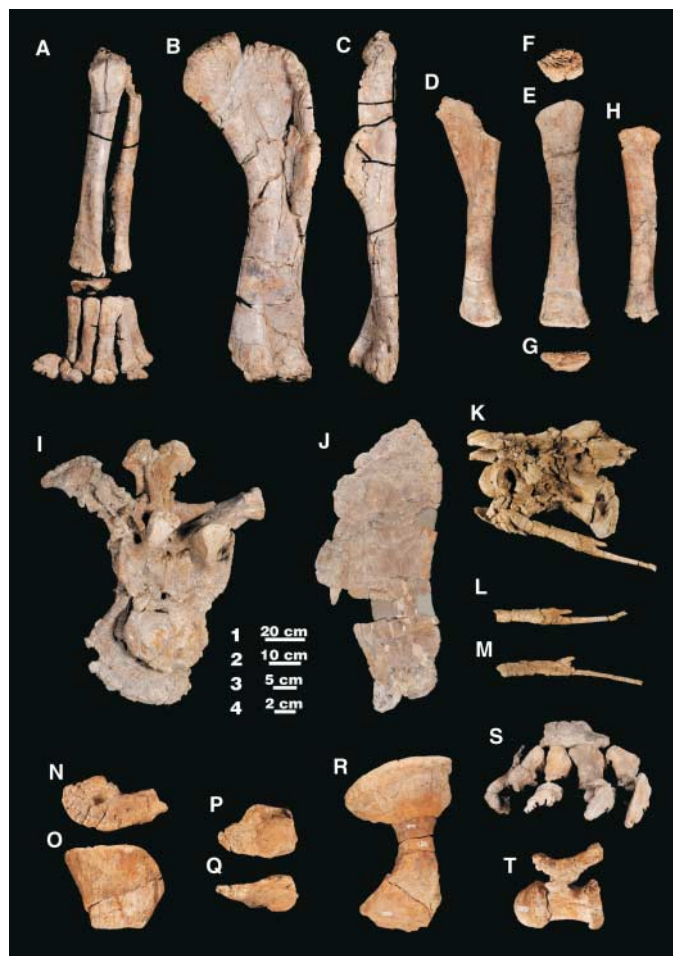


Fig. 1. Skeletal elements of *T. riodevensis* gen. et sp. nov.: left radius, ulna, carpal, and manus in cranial view (A); humerus in cranial (B) and lateral (C) views; left ulna in cranial view (D); left radius in medial (E), proximal (F), and distal (G) views; left fibula in medial view (H); middle dorsal vertebra in cranial view (I); left sternal plate in ventral view (J); cervical vertebra and rib in left lateral view (K); cervical rib in medial (L) and lateral (M) views; left tibia in proximal (N) and medial (O) views; left astragalus in proximal (P) and cranial (Q) views; metatarsal V in lateral view (R); right pes in cranial view (S); distal caudal vertebra in left lateral view (T). Scale bar 1 = 20 cm [(A) to (H), (N) to (Q), and (S)]; scale bar 2 = 10 cm [(I) to (M)]; scale bar 3 = 5 cm (R), and scale bar 4 = 2 cm (T).